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## SYNTHESIS OF 3'-(L-2-AMINO-3-PHENYLPROPYL)AMINO-3'-DEOXY-N<sup>6</sup>,N<sup>6</sup>-DIMETHYLADENOSINE -A NEW ANALOG OF PUROMYCIN

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The nucleoside antibiotic puromycin (5a) is a powerful inhibitor of protein synthesis which exhibits distinct anticancer activity.<sup>1</sup> Analogs of puromycin (5a) wherein the carbonyl group of amino acid moiety has been reduced to a methylene group are of interest because such a modification would not increase the size of the molecule but would enhance its resistance towards chemical and enzymic hydrolysis. Such a compound would also address an important problem of the mechanism of ribosomal protein synthesis - requirement for a carbonyl group (amide or ester) in the acceptor substrate.<sup>2,3</sup> Similar analogs of some peptides have found use in structure - activity studies.<sup>4,5</sup>

We now wish to report the synthesis of compound (5b) using a novel selective alkylation procedure (see Scheme 1). Thus, L-3-phenyl-2-phthalimido-1-propanol (1) obtained by a method used for the corresponding racemic compound<sup>6</sup> gave on reaction with trifluoromethanesulfonic anhydride in the presence of pyridine<sup>7</sup> in  $CH_2Cl_2$  trifluoromethanesulfonate 2. The interaction of 2 with puromycin aminonucleoside 3 using NaH in dimethylformamide (DMF) dioxane mixture afforded the N-phthaloyl derivative 4. The structure of 4 was confirmed by negative ninhydrin and NaIO<sub>4</sub>-benzidine<sup>8</sup> tests. The deblocking of 4 required the treatment with hydrazine in ethanol for 5 h at 40-50°C to afford the target compound 5b characterized as a hydrochloride salt. Product 5b gave a violet coloration with ninhydrin but it did not react with NaIO<sub>4</sub>-benzidine reagent.

Although alkylation of simple amines with esters of trifluoromethanesulfonic acid is documented<sup>9</sup> the present report is the first application to sterically hindered esters and complex polyfunctional amines. The presence of trifluoromethanesulfonyloxy group as an effective leaving group in 2 is essential for the success of the procedure. Thus, the corresponding methanesulfonyl derivative of 1 failed to react with aminonucleoside 3 under a variety of conditions.

Preliminary tests in a cell-free *E*. *coli* ribosomal system using N-acetyl-L-phenylalanyltRNA as a peptide donor showed that compound 5b at concentration  $1 \times 10^{-3}$ M is not capable of accepting N-acetyl-L-phenylalanyl residue from N-acetyl-L-phenylalanyl-tRNA and also it does not inhibit the puromycin reaction in the same system.<sup>10</sup> Further biological testing is in progress.

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## L-3-Phenyl-2-phthalimido-l-trifluoromethanesulfonyloxypropane-(2).<sup>11</sup>

A solution of compound 1 (ref. 6, 0.47 g, 1.7 mmol) and trifluoromethanesulfonic anhydride (0.47 g, 1.7 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 ml) was stirred 1 h at 0°C and 1 h at room temperature. The solvent was evaporated, the residue was washed with CHCl<sub>3</sub> (5 ml), the solution was filtered, the filtrate was evaporated and crude 2 was crystallized from 2-propanol, 0.5 g (73%), mp. 85-86°C; NMR (CDCl<sub>3</sub>)  $\delta$ , 3.25 (m, 2, CH<sub>2</sub> of benzyl), 4.71 (m, 1, CH), 5.01 (m, 2, CH<sub>2</sub>0), 7.19 (s, 5, C<sub>6</sub>H<sub>5</sub>), 7.73 (m, 4, phthaloyl); mass spectrum: m/e 413 (M<sup>+</sup>).

A suspension of aminonucleoside 3 (20 mg, 68 µmol) in DMF-dioxane (1:1, 4 ml) was stirred with NaH (2 mg, 83 µmol) for 30 min. at room temperature. Reagent 2 (28 mg, 68 µmol) was then added and the stirring continued for 17 h. The insoluble portion was filtered off, the filtrate was evaporated and the residue was chromatographed on a loose layer of silica gel<sup>12</sup> in  $CH_2Cl_2$ - $CH_3OH$  (93:7). The faster moving band was eluted, the eluate was evaporated and crude 4 was crystallized from 2-propanol or ethyl acetate, 21 mg (55%), mp. 202-203°C;  $UV_{max}^{EtOH}$  267 nm ( $\epsilon$  17 300), shoulder 283 nm ( $\epsilon$  15 600); NMR ( $CD_3SOCD_3 + D_2O$ )  $\delta$ , 3.50 (s, 6, ( $CH_3$ )<sub>2</sub>N), 6.11 (d, J<sub>1</sub>, 2<sup>1</sup> 3 Hz, 1, H<sub>1</sub>), 7.18 (s, 5,  $C_6H_5$ ), 7.73 (m, 4, phthaloyl), 8.31 (s, 1, H<sub>2</sub>), 8.46 (s, 1, H<sub>8</sub>); mass spectrum: m/e 557 (M<sup>+</sup>). [ $\alpha$ ]  $_D^{24}$  - 6.2° (c 0.15, ethanol).

## <u>3'-(L-2-Amino-3-pheny1propy1)amino-3'-deoxy-N<sup>6</sup>,N<sup>6</sup>-dimethy1adenosine (5b).</u>

A mixture of compound 4 (20 mg, 36  $\mu$ mol), hydrazine hydrate (1 ml) and ethanol (4 ml) was heated for 5 h at 40°-50°C. After evaporation of solvents the residue was chromatographed on a loose layer of silica gel<sup>12</sup> in CH<sub>2</sub>Cl<sub>2</sub>-CH<sub>3</sub>OH (4:1). The major UV absorbing band was eluted, the eluate was evaporated, product was dissolved in methanol (20 ml), the solution was saturated with HCl and then it was kept at -10°C for 3 days. The insoluble portion was filtered off, the filtrate was evaporated and the residue was crystallized from 80% ethanol, 11 mg (66%, hydrochloride); mp. 223-227°C; UV  $_{max}^{EtOH}$  275 nm ( $\epsilon$  13 600); NMR (CD<sub>3</sub>SOCD<sub>3</sub> + D<sub>2</sub>O)  $\delta$ , 3.46 (s, 6, (CH<sub>3</sub>)<sub>2</sub>N), 5.99 (d, J<sub>11,21</sub> 3 Hz, 1, H<sub>11</sub>), 7.25 (s, 5, C<sub>6</sub>H<sub>5</sub>), 8.21 (s, 1, H<sub>2</sub>), 8.40 (s, 1, H<sub>8</sub>). [ $\alpha$ ]  $_{2}^{2}$  - 9.8° (c 0.12, H<sub>2</sub>O).

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## REFERENCES AND FOOTNOTES

- 1. R.J. Suhadolnik, Nucleoside Antibiotics, p. 3., Wiley-Interscience, New York, 1970.
- 2. R.J. Harris and R.H. Symons, Bioorg. Chem. 2, 286 (1973).
- 3. A.A. Krayevsky, M.K. Kukhanova and B.P. Gottikh, Nucleic Acids Res. 2, 2223 (1975).
- 4. E. Atherton, H.D. Law, S. Moore, D.F. Elliott and R. Wade, J. Chem. Soc. (C) 3393 (1971).
- 5. K.-F. Fok and A. Yankeelov, Jr., Biochem. Biophys. Res. Commun. 74, 273 (1977).
- 6. S. Yamada, K. Koga and H. Matsuo, Chem. Pharm. Bull. 11, 1140 (1963).
- 7. C.D. Beard, K. Baum and V. Grakauskas, J. Org. Chem. 38, 3673 (1973).

- 8. M. Viscontini, D. Hoch and P. Karrer, Helv. Chim. Acta 38, 642 (1955).
- 9. R.D. Howells and J.D. McCown, Chem. Rev. 77, 69 (1977).
- 10. P. Bhuta, unpublished results.
- 11. All new compounds gave satisfactory C, H, N analyses and they were homogeneous on TLC.<sup>12</sup> NMR spectra were measured on FT spectrometer JEOL FX 100 and mass spectra on JEOL JMS 01SG-2. Tetramethylsilane was used as an internal standard with CDCl<sub>3</sub> and sodium 2, 2-dimethyl-2-silapentane-5-sulfonate with CD<sub>3</sub>SOCD<sub>3</sub>. Evaporations were carried out on a rotary evaporator at 30°C. Melting points were determined on a Thomas-Hoover capillary melting point apparatus and are not corrected.
- 12. J. Žemlička and J. Owens, J. Org. Chem. 42, 517 (1977).

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